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Conditional colored and colorless alleles at the C₁ locus.

Besides the dominant colored allele, \underline{C}_1 , and inhibitor, \underline{C}_1^I , recessive colorless, \underline{c}_1 , can be differentiated into two allelic forms: \underline{c}^p (p for positive), the conditional colored which becomes colored in the light during germination (MGCNL 44:153, 1970), and \underline{c}^n (n for negative), the colorless which fails to give color at any time. Their dominance relationship is $\underline{c}^I > \underline{c} > \underline{c}^p > \underline{c}^n$. It has been found that \underline{c} testers derived from W22 and K55 contain conditional colored \underline{c}^p alleles while most other \underline{c} tester stocks and KYS contain colorless \underline{c}^n alleles. Four point linkage data have been obtained to support the differentiation of \underline{c} alleles into two forms (Table 1).

Some properties of this conditional colored \underline{c}^P allele have been observed. Light is necessary for pigment formation in \underline{c}^P tissue. Peeling off the husk to expose \underline{c}^P tissue to the light does not induce pigment formation during normal seed development. However, \underline{c}^P \underline{vp}_1 kernels on the cob can synthesize pigments (although pigments are reduced) if the husk is removed. Thus, light and germination are two critical conditions for anthocyanin formation in \underline{c}^P tissue. The \underline{c}^P allele has many properties in common with \underline{c} . It is inhibited by \underline{c}^I and shows mottled expression with $\underline{R} \ \underline{r} \ \underline{r}$. It also requires \underline{A}_1 , \underline{A}_2 , \underline{C}_2 , \underline{Bz}_1 , and \underline{Bz}_2 to be present in the dominant form for pigment formation. Unlike \underline{C} , \underline{c}^P seeds from $\underline{c}^P/\underline{c}^P$ \underline{c}^P segregating ears sometimes show a wide range of color variation, from very light to very intense. The variation is not heritable. It is possible that the variation is due to dosage effect at the \underline{c} locus, to background factors, to varying physiology and vigor of germinating seeds, or to environmental factors.

The relative concentration of cyanidin to pelargonidin (obtained by acid hydrolysis of pigments) in \underline{c}^p germinated seeds is much lower than that in normally pigmented \underline{c} seeds (Kirby & Styles, Can. J. Genet. Cytol. 12:934, 1970). We observed similar results (lower cy/pg ratio) in \underline{c}^p

Table 1 Four point linkage data for $\frac{+ cP + +}{yg c^n sh wx} \times yg c^n sh wx$

Test cross	Parental	Rl	R2	R3	R1 & R2	R1 & R3	R2 & R3	R1, R2 & R3	Total
c ^p (W22)	1731 1580 3311	1094 19 . 59%	168 3.01%	954 17.08%	19 0.34%	31 0.56%	2	5 0.0%	5584
c ^p (K55)	963 892 1855	233 279 512 16.20%	68 79 147 4.65%	339 272 611 19.33%	1 5 6 0.19%	5 5 8 13 0.41%	4 5	4 4 8 0.25%	3161

tissue in K55 do not show a decrease in cy/pg ratio, compared with that of $\underline{\mathbf{C}}$ tissue (Table 2). Within W22 background, \mathbf{F}_1 ($\underline{\mathbf{C}}$ $\underline{\mathbf{C}}$ x $\underline{\mathbf{c}}^p\underline{\mathbf{c}}^p$) and \mathbf{F}_2 $\underline{\mathbf{C}}$ --

Table 2
Relative concentration of anthocyanidins from \underline{C} and \underline{c}^p tissues in different backgrounds (average 0.D. of 4 replicates).

Background		W22		K55		
Anthocyanidin	су	pg	cy/pg	су	pg	cy/pg
Genotype						
A C R	2.64	0.17	15.53	1.60	0.39	4.10
A c ^p R	1.69	0.92	1.84	0.64	0.14	4.57
$F_1(C C \times c^p c^p)$	1.54	0.10	15.40			
	1.64	0.14	11.71			
F ₂ C F ₂ c ^p c ^p	0.59	0.32	1.84*			

^{*}two replicates cy = cyanidin, pg = pelargonidin

seeds have a cy/pg ratio similar to that of the \underline{C} \underline{C} parent, while F_2 conditional colored $\underline{c}^p\underline{c}^p$ seeds have a ratio similar to that of the $\underline{c}^p\underline{c}^p$ parent. The data favor the hypothesis that physiological conditions of germination in W22 result in the increase in pelargonidin. Anthocyanidin constitutions of F_1 and F_2 between K55 \underline{c}^p and W22 \underline{c}^p have also been studied (Table 3).

Table 3 Relative concentration of anthocyanidins of $F_1 c^p$ (W22) $x c^p$ (K55), F_2 , and backcrosses (average O.D. of 4 replicates)

Anthocyanidin		Relative O.D.	
family	су	pg	cy/pg
F	1.64	0.70	2.35
F ₂	0.71	0.38	1.87
$F_1 \times \underline{c}^p$ (W22)	1.50	0.74	2.03
F_2 $F_1 \times \underline{c}^p$ (W22) $F_1 \times \underline{c}^p$ (K55)	1.12	0.44	2 . 54

The data indicate that the two \underline{c}^p alleles present in W22 and K55 \underline{c} testers have essentially the same effects. The materials used for K55 \underline{c}^p were from very weak inbred ears, and this may be the reason for the higher cy/pg ratio observed in K55 \underline{c}^p tissue. There seems to be a general trend such that the more vigorous the germinating seed, the lower the cy/pg ratio.

It is clear that there are two allelic forms present in \underline{c} testers: \underline{c}^p , the conditional colored, and \underline{c}^n , the colorless. Light and germination are two required conditions for anthocyanin formation in the \underline{c}^p tissue. The pigments formed in \underline{c}^p tissue have a lower cyanidin/pelargonidin ratio than that in normally pigmented \underline{c} tissue.

Shu-mei Chen

2. An unsuccessful search for mutations affecting anthocyanin distribution.

Large populations of one inbred line, Ky 27, grown in isolation, have been observed closely for mutations of factors controlling anthocyanin distribution. This inbred is $\underline{A} \ \underline{C} \ \underline{r}^{r}$ and has purple plumule; a search was conducted in the plants and in the seeds produced on them for pigment formation in new locations -- for example, in culm, husk, glume bar, and aleurone tissue. Among more than 10,000 plants studied, no distinctive plants or sectors were found; among 3.62 x 106 kernels, several colored contaminations were identified, but no valid mutations were found, either as whole-kernel exceptions or as sectors down to the limit of naked-eye resolution. Considering that any single mutation-competent locus (for example, \underline{r}^{r}) is present in the aleurone in 3 doses (i.e., 10.86 x 10⁶ chromosomes entering into the triple fusion), and that the twofold observation protocol used here should identify events through at least the first ten divisions (i.e., 103 sites for minimum detectable mitotic events; 10⁶ mitoses per kernel), no mutation to anthocyanin synthetic capacity was found in around 10¹³ mitotic replications. This observing load was lightened by the help of Paul Bolen, Shu-mei Chen, John Cousins, Kenneth Hall, Henry Lee, Marion Murray, Donald Smith, Jean Spengel, and Charles Williamson.