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1. Chemical treatment of seeds heterozygous for Wd and Yg2.

In this experiment Yg₂ yg₂ and Wd wd heterozygous seeds were soaked in the dilute chemicals which had been found or are suspected to be mutagenic in lower and in higher organisms, frequently stirred, and rinsed after 8 hours of treatment at 23° C. The loss of the dominant genes or the absence of their function was observed phenotypically as yellow green or albino sectors on green background. The Wd wd heterozygote was produced by crossing a wd wd male containing Yg Wd Cl as an independent ring (obtained from Dr. G. Y. Kikudome) on a stock having normal chromosome 9 with the genes Yg Wd C. The role of the ring is to maintain the wd wd homozygous deficient stock. The ring is frequently lost during division resulting in albino stripes and a high proportion of gametes that are wd. For treatment the fully colored seeds were used since they probably lacked the ring. However it should be noted that rings which may have lost a segment of the ring carrying Cl during division, but retained either Wd, Yg or both are not detectible by this technique.

The following can be concluded from the table:

- 1. Ethyl methanesulfonate (EMS) and Diethyl Sulfate produce sectors in both leaf and shoot, the frequency being highest in the first leaf and linearly decreasing in subsequent leaves. The sectors are the smallest on the first leaves gradually increasing in size on subsequent leaves.
- 2. The mutagenic effect of Glycidol, Epichlorohydrin, Colchicine, although approaching significance, remains uncertain.
- 3. The frequency of sectors is of the same magnitude in both heterozygotes. This observation supports the assumption that colored seeds were <u>Wd</u> wd heterozygotes since, had the functional rings devoid of C^I carrying the genes <u>Wd</u>, <u>Yg</u> or both been frequently transmitted, the heterozygote seedlings could not be uniformly sectored.
- formed in excess since a break between the <u>Wd</u> locus and the centromere should give albino sectors and should be more frequent than breaks between <u>Wd</u> and <u>Yg</u>. On the contrary the <u>Wd</u> wd heterozygotes produced an excess of yellow green sectors. This observation indicates that EMS either preferentially breaks the chromosome toward the end, or produces a high proportion of mutations as opposed to chromosome breaks. Present data do not permit proper distinction between these possibilities.

Chemical	Conc. M/lit	Lo- cus	No. of seeds treated	Sur- viving seed- lings (%)	ter treat-	\sim per leaf (f) and size ² (s)													
						lst %	leaf fs	2nd %	leaf fs	3d %	leaf fs		leaf fs		leaf fs	6th %	leaf fs		leaf fs
Ethyl Methane- sulfonate	0.05 0.05 0.10 0.10 0.20	Yg Wd Yg Wd	101 30 50 30 20	100.0 100.0 92.0 96.6 5.0	40.4 39.2 35.0 31.5 8.0	100 100 100 100 100	6a 7a 7a	100 100 100 100 100	5a 6a 6a	100		90 100	Lab Lab Lab Lab	** 9: ** 9: **100	3bc 3bc	**56 **29 **73 **67	2bc 3abc	*71	2bc 2bc 2bcd 2bcd
Diethyl Sulfate	0.05 0.10	Yg Yg	100 49	19.0 0.0	13.2	74 	5a 	74 	5a 	82	Ца 		4ab	90) 3ab	33 	2ab		
Glycidol	0.05 0.10	Yg Yg	100 42	92.0 89.7	38.0 33.9	17 100	lc lc	-		-	 	•))	•		-	
Epichloro- hydrin	0.05 0.05 0.10 0.15 0.50	Yg Wd Yg Yg Yg	100 30 49 30 20	69.0 100.0 59.2 0.0 20.0	34.9 36.4 31.2 	7	lc lc lc	0		C)))	*0 0	la	* ·	0 la 0 0 5 lb	*10 * 0 * 0		*10 * 0 * 0	
Colchicine	0.01 0.01 0.02 0.02	Yg Wd Yg Wd	100 47 50 49	92.0 100.0 92.0 91.8	42.8 36.1 39.9 34.8	7 13		7 13	•	7 13) ца 1 ца 3 5а 1 ца	7 9	_		1 1b 5 1b 2 1a 2 1b	2	1b 1b	0	
Control		Yg Wd	200 70	98.4 100.0	40.5 38.6	-)	-) 1 lb	_)))		0 0 la	-))	_))

(See next page for footnotes.)

 $\frac{1}{\text{Frequency index:}}$ (7) = 101<, (6) = 51-100, (5) = 11-50, (1) = $\frac{1}{1-10}$, (2) = 1-2, (1) = 1 sector per leaf respectively.

²Sector size (a) -- 1 mm wide, length from 1 mm to 3/4 length of leaf; (b) -- < 1.5 mm; (c) -- 1.5-4 mm; (d) -- 4-10 mm wide, and extending from base to tip or to margin of leaf.

*Random sample (10-20% of surviving seedlings). **Random sample (50% of surviving seedlings).

G. Ficsor

2. Transposition of mutability between components of the A1 locus.

In studies of two separate cases of mutability arising from potentially compound alleles of the \underline{A}_1 locus mutant types have occurred which suggest transposition of the factor responsible for mutability from one component to the other.

The first of these (MNL 30:101) originated from \underline{A}^b and appeared to be composed of a stable $\underline{\alpha}$ and a mutable recessive $\underline{\beta}$ component ($\underline{\beta}^m$). The instability is controlled by one or more separate and as yet unidentified factors. The $\underline{\alpha}$ $\underline{\beta}^m$ complex usually behaves just as would be expected on the basis of its structure. The $\underline{\alpha}$ component may be removed by crossing over to produce a colorless mutable allele $\underline{\beta}^m$, the $\underline{\beta}^m$ may change to recessive stable to produce a pale stable allele, the $\underline{\beta}^m$ may change to dominant stable and thus produce a reconstituted $\underline{A}^{\underline{b}}$ ($\underline{\alpha}$ $\underline{\beta}$) or the $\underline{\beta}$ component may change its state of mutability to produce a more or less unstable allele.

An occasional exception is found in the occurrence of cases where the mutability appears to have transferred to the α component leaving the β component in a recessive and much less mutable condition. Seeds of such a type are colorless with many pale and a few full colored sectors.

The second case originated several years ago in L. J. Stadler's cultures from the standard a^p allele. The new allele designated a^{pm} arose from a less mutable allele designated a^{px} which in turn arose from a^p. Because of its phenotypic expression and its failure to respond to attempts to subdivide it by crossing over the a^p allele was considered to be a single unit allele similar to the a^r component of A^p Peru. However when one considers the behavior of its descendant allele a^{pm} one is led to conclude otherwise.

Regularly and without the need of any known mutator factor \underline{a}^{pm} which has pale aleurone, red brown plant and dominant brown pericarp color (\underline{a} phenotype) changes to \underline{A}^r which has a purple seed, purple plant and recessive red pericarp color ($\underline{\beta}$ phenotype) or to \underline{A}^{br} which has purple seed, purple plant and recessive brown pericarp color. Thus the mutants produced fail to fit into a linear series expected if a one-component locus were involved nor do they fit a two-component locus since the mutants obtained require that both components change in opposite directions at the same time. This seems quite illogical until one considers the possibility that such a dual change can occur if one component gives up something at the same time that the other gains